

## Chemical reactivity of cisplatin bound to human plasma proteins

L. Hegedüs<sup>1</sup>, W. J. F. van der Vijgh<sup>2</sup>, I. Klein<sup>2</sup>, S. Kerpel-Fronius<sup>1</sup>, and H. M. Pinedo<sup>2</sup>

<sup>1</sup> National Institute of Oncology, Budapest, Hungary

<sup>2</sup> Academic Hospital, Free University, Amsterdam, The Netherlands

**Summary.** Human plasma was incubated with cisplatin over 24 h. Ultrafilterable platinum and platinum reactive with DDTC were determined at regular time intervals during incubation. At each time point more platinum reacted with sodium *N,N'*-diethyldithiocarbamate (DDTC) than was available as ultrafilterable platinum. At 24 h 70% of total platinum (10% ultrafilterable platinum and 60% protein-bound platinum) reacted with DDTC. This means that cisplatin bound to plasma proteins can – at least in part – still react with strong nucleophiles.

### Introduction

Binding of cisplatin to plasma proteins has been studied by measuring concentrations of ultrafilterable platinum [1, 5]. Only a few papers have been published about the biological activity [4, 6], and nothing is known about the chemical reactivity of protein-bound platinum. The present experiment has been carried out to study the chemical reactivity of protein-bound platinum with sodium diethyldithiocarbamate (DDTC). DDTC was chosen because of its strong nucleophilic properties and its possible clinical application for reduction of cisplatin-induced nephrotoxicity [3].

### Materials and methods

Cisplatin was kindly supplied by Bristol Myers (Weesp, The Netherlands). Sodium DDTC (analytical grade) was purchased from Merck (Amsterdam, The Netherlands). The PtDDTC<sub>2</sub> complex was synthesized by the procedure of Bannister et al. [2]. Binding of cisplatin to human plasma proteins was studied *in vitro* by incubating 50  $\mu$ M cisplatin in plasma at 37° C for 24 h. At 0, 0.5, 1, 2, 4, 6, 8 and 24 h after the start of incubation 1-ml aliquots were taken for ultrafiltration and for reaction with sodium DDTC. Ultrafiltration was performed in Amicon MPS-1 micropartition systems (Amicon, Oosterhout, The Netherlands) containing YMT membranes (cut-off 30000 daltons) at 2000 g in a bench-top centrifuge with a fixed angle rotor [7]. The first 8-min fraction was collected for the determination of ultrafilterable platinum (sol. A). DDTC-reactive platinum was determined by adding 100  $\mu$ l 2.5.10<sup>-2</sup> M DDTC in wa-

ter (which means a 500-fold excess with respect to platinum present in plasma) to a 1-ml aliquot of plasma, followed by incubation at 37° C for 4 h. The mixture was fractionated by a Sep Pak C-18 cartridge (Waters, Etten-Leur, The Netherlands): the eluate was obtained immediately after introduction of the complete reaction mixture in plasma (Sol. B) and the eluate, after purging of the cartridge with 50 ml methanol containing the reaction product PtDDTC<sub>2</sub>. The methanol solution was evaporated under vacuum and the residue redissolved in 2 ml ethanol (Sol. C). The platinum compounds retained on the Sep Pak column were removed by digesting the packing material with 3 ml 65% HNO<sub>3</sub> at 170° C in a polytetrafluoroethylene (PTFE) bomb for 2 h. The remaining liquid was evaporated at 120° C under a stream of air after addition of 5 mg NaCl. The residue was dissolved in 1 ml 0.2 N HCl-0.15 M NaCl (Sol. D). Platinum concentrations in solutions A–D were determined by atomic absorption spectrometry as described before [8]. Standard solutions of cisplatin were prepared in the same matrix as the samples.

### Results and discussion

The amounts of platinum detected in the four above-mentioned solutions are summarized in Table 1. The amounts of platinum present in solution A represent ultrafilterable platinum. The observed biphasic decrease of the platinum concentration with time corresponded with the results of previous studies [1, 2, 4, 6]. Platinum concentrations in solution B represent platinum irreversibly bound to proteins. This amount increased with time. However, if all platinum bound to plasma proteins is actually irreversibly bound then the amounts present in the solutions A and B together should total 100%. The lower values observed indicate that some of the protein-bound platinum species can be removed by the strongly nucleophile DDTC. This is in agreement with the higher platinum concentrations in solution C, representing Pt-DDTC adducts, than in solution A. Nevertheless, two peculiarities are observed: (a) the amounts present in solutions B and C together do not add up to 100%; and (b) the amounts of platinum present in solution C were expected to decrease continuously with time, but the amounts at the two latest time points were higher than the preceding values. In an attempt to find an explanation for these observations the amount of platinum retained on the column was measured at various time points. The amounts present in solution D did indeed complete the 100% platinum balance when added to the amounts

Offprint requests to: W. J. F. van der Vijgh, Academisch Ziekenhuis Vrije Universiteit, De Boelelaan 1117, 1081 HV Amsterdam, The Netherlands

**Table 1.** Platinum, expressed as percentages of total platinum (50  $\mu$ M) in plasma, present in plasma ultrafiltrate (Sol.A), plasma eluate from the Sep Pak cartridge (Sol.B), methanol eluate from the Sep Pak cartridge (Sol.C) and final solution after reaction of packing material with nitric acid (Sol.D)

Time (h)	Sol.A	Sol.B	Sol.C	Sol.D
0.5	70.2 $\pm$ 1.0	5.1 $\pm$ 0.1	90.5 $\pm$ 8.6	ND
1	61.3 $\pm$ 0.8	7.4 $\pm$ 0.2	81.4 $\pm$ 7.8	13.0 $\pm$ 0.2
2	46.6 $\pm$ 0.4	10.8 $\pm$ 0.2	59.0 $\pm$ 5.0	ND
4	30.4 $\pm$ 0.2	15.6 $\pm$ 0.2	49.2 $\pm$ 4.2	40.2 $\pm$ 1.0
6	21.5 $\pm$ 0.1	19.2 $\pm$ 0.2	43.7 $\pm$ 3.6	ND
8	19.3 $\pm$ 0.1	21.1 $\pm$ 0.2	56.5 $\pm$ 4.8	12.8 $\pm$ 0.2
24	11.3 $\pm$ 0.1	28.6 $\pm$ 0.2	68.4 $\pm$ 6.2	1.5 $\pm$ 0.1

ND, not determined

present in solutions B and C. Furthermore, the observed increase and subsequent decrease in the amounts with time are complementary to the values observed in solution C. Because of their retention behaviour it is thought that the compounds present in solution D originally represented platinum bound to protein as well as DDTC. If so, it can be understood that formation of such compounds will lead to an increase of platinum in solution D with time, but also that at a certain time, when only small amounts of ultrafilterable platinum are left, replacement of DDTC by a second protein leads to a shift of platinum from solution D to solution C, as observed at 8 and 24 h after the start of incubation. It may be concluded that although dialysis experiments have suggested that non-ultrafilterable platinum species are irreversibly bound to plasma proteins [5], our observations indicate that at least some protein-bound

platinum can react with strongly nucleophile substances, for example, which has implications for the fate of cisplatin in vivo and the use of rescue therapies.

## References

1. Bannister SJ, Chang Y, Sternson LA, Repta AJ (1978) Atomic absorption spectrophotometry of free circulating platinum species in plasma derived from *cis*-dichlorodiammineplatinum (II). *Clin Chem* 24: 877
2. Bannister SJ, Sternson LA, Repta AJ (1979) Urine analysis of platinum species derived from *cDDP* by HPLC following derivatization with DDTC. *J Chromatogr* 173: 333
3. Gale GL, Atkins LM, Walker EM (1982) Further evaluation of diethyldithiocarbamate as an antagonist of cisplatin toxicity. *Ann Clin Lab Sci* 12: 345
4. Gullo JJ, Litterst CL, Maguire PJ, Sikic BI, Hoth DF, Woolley PV (1980) Pharmacokinetics and protein binding of *cis*-dichlorodiammineplatinum (II) administered as a one hour or as a twenty hours infusion. *Cancer Chemother Pharmacol* 5: 21
5. Repta AJ, Long DF (1980) Reaction of cisplatin with human plasma and plasma fractions: In: Prestayko AW, Crooke ST, Carter SK (eds) *Cisplatin, Current status and new developments*. Academic Press London, p 285
6. Takahashi K, Seki T, Nishikawa K, Minamide S, Iwabuchi M, Ono M, Nagamine S, Horinishi H (1985) Antitumor activity and toxicity of serum protein-bound platinum formed from cisplatin. *Gann* 76: 68
7. Van der Vijgh WJF, Klein I (1986) Protein binding of five platinum compounds. Comparison of two ultrafiltration systems. *Cancer Chemother Pharmacol* 10: 129
8. Vermorken JB, van der Vijgh WJF, Klein I, Hart AAM, Gall HE, Pinedo HM (1984) Pharmacokinetics of free and total platinum species after short-term infusion of Cisplatin. *Cancer Treat Rep* 68: 505

Received August 12, 1987/Accepted May 13, 1987